The evolution of neuronal circuits underlying species-specific behavior

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The nervous system is evolutionarily conservative compared to the peripheral appendages that it controls. However, speciesspecific behaviors may have arisen from very small changes in neuronal circuits. In particular, changes in neuromodulatory systems may allow multifunctional circuits to produce different sets of behaviors in closely related species. Recently, it was demonstrated that even species differences in complex social behavior may be attributed to a change in the promoter region of a single gene regulating a neuromodulatory action.

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Current Opinion in Neurobiology 1999, 9:628-633

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Abbreviations

FMRFamide	Phe-Met-Arg-Phe-amide
GABA	γ-aminobutyric acid
GPR	gastropyloric receptor
JAR	jamming avoidance response
NMDA	N-methyl-D-aspartate
STG	stomatogastric ganglion

Introduction

Neuronal basis of species-specific behavior

Natural selection acts on many aspects of organisms, from their appearance to their molecular makeup. Evolutionary pressures also shape the behavior of organisms, producing species-specific behaviors. Although it is a difficult task to determine the neuronal basis for complicated behavior even in a single species, insights into neuronal circuit design can be gained by comparing closely related species with different behaviors. The variations seen between the species' nervous systems could serve as natural experiments for probing the role of those structures in the production of behavior [1]. Furthermore, determining the extent of the differences between the nervous systems of two species with different behaviors can give an indication of how finely tuned nervous systems must be in order to generate appropriate behaviors.

Numerous examples exist showing that neuronal differences can cause changes in behavior between closely related species [2–4]. For example, two closely related species of deer, the white-tailed deer (*Odocoileus verginianus*) and the mule deer (*O. hemionus*) use different gaits when alarmed. The white-tailed deer gallop, whereas the mule deer stot. It can be inferred that this species difference in behavior is caused by genetic differences in the nervous systems of the two species because hybrid offspring produce an intermediate behavior: they bound when alarmed [2]. Recently, the neural basis for speciesspecific behavior was dramatically demonstrated by transplanting vocalization-related brain tissue from a Japanese quail to a chicken embryo, and transforming the vocalizations from one species to the other [5^{••}]. Progressive changes in behavior can also be traced in certain lineages in which new behaviors evolved as a series of elaborations of pre-existing specialized behaviors [6[•],7,8,9[•]].

Mechanisms underlying species differences in behavior

Numerous factors might contribute to producing a difference in the behaviors of closely related species. First, the species could differ in their acquisition of sensory signals. There are many examples of elaboration or loss of sensory structures in related species [10,11]. Second, the central neuronal networks of two related species might process information differently. This could involve changes in the number or properties of cells and synapses in the central nervous system or in their pattern of connectivity [12–15]. Finally, the peripheral structures might be modified to respond differently to central commands. Changes in the relative size, position, or number of muscles can produce qualitatively different movements from the same neuronal motor pattern [9•,16,17•].

It has been suggested that because peripheral structures are subject to fewer constraints, they might be more likely than central circuits to exhibit phylogenetic differences ([12–14,18]; see, however, [19]). However, general properties of the nervous system and specific developmental mechanisms serve to keep the nervous system matched to a changing periphery. For example, a change in the size of sensory or motor structures will induce a corresponding change in the number of central neurons that serve that structure [20,21]. In addition, some changes in sensory input can be automatically accepted by existing neuronal circuits without necessarily inducing any central changes [22^{••}]. In this review, we will focus primarily on how cellular changes to central neuronal networks may mediate the evolution of species-specific behaviors.

Are neurons conserved through evolution?

The basic organization of nervous systems tends to be highly conservative. Members of species belonging to the same genus often have almost indistinguishable nervous systems. In invertebrates, where individual neurons can be unambiguously identified from animal to animal within a species, homologous neurons can be identified in disparate members of a taxon such as Insecta [23] and even across phyla [24[•]]. Of course, homologous neurons can exhibit certain morphological modifications that are taxon specific [25].

Homologous neurons in disparate species also can be identified by neurochemical criteria. For example, homologous serotonin-immunoreactive neurons can be identified in highly divergent species within the class of gastropod molluscs [26]. Similarly, in the class Crustacea, serotonergic neurons that have been identified in highly derived decapod species such lobsters and crayfish [27] are also found in primitive anaspid species such as *Anaspides tasmaniae* [28]. Although there are important examples of prominent neurons that have been lost during evolution in some members of a lineage [29], our impression is that identified neurons are often conserved during evolution.

The most common difference seen in closely related species is a change in the number of cells of a particular type [14,30,31[•],32,33,34[•]]. The organizational features of some neuronal structures, such as the laminar organization of neuronal cell types (e.g. as in the cerebral cortex, the retina, and certain nuclei), provide a simple mechanism for addition or subtraction of identical units, leading to computationally important changes in neuronal number [35[•],36].

Is circuit organization conserved through evolution?

Not only can individual homologous neurons be identified across species within a phylum but, as might be expected, entire circuits are conserved, even across phylogenetic orders. For example, the basic organization of the central pattern generators for locomotion is conserved in a variety of vertebrate spinal cords, including the spinal cord of lamprey, larval *Xenopus*, and neonatal rat [37[•]]. However, differences in the detailed cellular properties of spinal neurons and the amount of excitatory input that they receive have been noted, even between closely related species [38[•],39]. Similarly, many of the same connections are found between homologous neurons underlying escape swimming in two molluscs of the subclass Opisthobranchia: the nudibranch *Tritonia diomedea* and the notaspid *Pleurobranchaea californica* [40[•]].

The complexity of the nervous system makes it difficult to determine the extent of phylogenetic differences in neuronal circuits. Therefore, work on the numerically simple crustacean stomatogastric ganglion (STG), which contains just 30 neurons, has been enlightening. The neuronal circuitry of the STG has been remarkably well preserved over at least 350 million years of evolutionary divergence, despite radical changes in the peripheral structures that it controls [15,41**,42-44]. Most of the neurons in the pyloric network are identifiable in all species, though the numbers of some of the cell types can vary. The overall synaptic circuitry is similar, but there are differences in the relative strength of particular synaptic connections and the amount of electrical coupling between neurons in different species [41**,43,44,45*]. In some species, the intrinsic properties of

individual neurons have diverged, causing similar circuits to produce different motor patterns [42,43,46]. Most of the identified neurons have retained their transmitter phenotype across species; however, in one more distantly related species, the transmitter of two neurons is different but the postsynaptic ionic response to those neurons has remained the same as in the other species [47].

Constraints attributable to multifunctional networks

One possible reason that neural networks are so well preserved across species is that neuronal circuits act as generalists rather than specialists: a single motor circuit can produce a variety of different motor patterns under different circumstances [48–53] or at different times during the development of the animal [54-57]. These different behavioral outputs depend upon the actions of neuromodulatory inputs, as well as sensory feedback. Modulatory inputs can reconfigure networks by altering the strength of synapses and changing the intrinsic firing properties of the component neurons through the release of substances such as amines and neuropeptides [52,58]. In addition, modulatory inputs can cause neurons to switch allegiance from one network to another and, in some cases, can cause the fusion of multiple independent networks into a single conjoint network controlling a complex behavior [48,59-63]. This flexibility in the output of neuronal networks has two evolutionary consequences. First, there is no need to evolve a completely new circuit to produce a new behavior. Second, the fact that a network must play roles in many different behaviors or at different developmental stages may constrain it from being altered because changes in the network that would be advantageous for one behavior might be disastrous for another.

Changes in the input to a circuit

If neuronal circuitry evolves more slowly than behavior, then perhaps natural selection can alter the range of behaviors produced by a circuit by changing its inputs or by changing how it handles those inputs. Furthermore, it may be more parsimonious to alter the inputs to a circuit than to change the connectivity within a circuit itself. The electrosensory system in South American electric fish provides an excellent example of behavioral differences arising from differences in the inputs to similar circuits. The pacemaker nucleus underlying the jamming avoidance response of the two closely related South American electric fish genera, Eigenmannia and Apteronotus, is similar. However, the nucleus receives different sets of inputs from other brain areas that cause a different behavioral response in the two species [64,65[•]]. Variations in the inputs to circuits have been shown to play a role in the expression of species-specific behavior in tadpoles [66-68], and species differences in the aminergic input to the mammalian cerebellum have also been reported [69]. In addition, many phylogenetic variations have been observed in the neuromodulatory inputs to the STG [15]. For example, serotonergic innervation of the STG is

provided by a set of muscle receptor neurons that have been identified in at least eleven species from six decapod crustacean infraorders. There are species and lineage differences in the number of these neurons (varying from one to four pairs), the muscles that they innervate, and their apparent peptide co-transmitters (e.g. allatostatin-, cholecystokinin-, and FMRFamide-like immunoreactivity) [70•,71–73]. Furthermore, in the spiny lobsters, the neurons are present, but they do not contain serotonin [74]. Instead, serotonin is thought to be delivered as a circulating neurohormone. Species differences in the mode of delivery of a neuromodulatory substance, including release from different neurons with different co-transmitters and targets in the network, could dramatically alter the role that the compound plays in the production of behavior [71,75,76,77•].

Changes in responses and receptors

The response of particular neurons or brain areas can change even if the transmitters involved stay the same [53,74,78]. For example, serotonin is known to have two effects on sensory neurons in the mollusc *Aplysia californica* that play a role in non-associative learning: it increases their excitability and causes spike broadening. However, in other related species, the spike broadening response is absent [79]. This may be attributable to differences in the types of serotonin receptors on these identified neurons or the coupling of the receptors to their second-messenger systems. In one species, both the spike broadening and excitability responses are absent, and this species also lacks dishabituation and long-term sensitization in the tail withdrawal reflex, which are thought to be mediated by serotonin in *Aplysia* [80].

Changes in receptor type or distribution can lead to marked changes in behavior. A particularly striking example of this is seen in two species of voles that differ in their affiliative behavior. The prairie vole (Microtus ochrogaster) forms monogamous pair bonds, whereas the montane vole (Microtus montanus) is solitary and does not show a preference for former mates. Two peptide transmitters, oxytocin in the female and vasopressin in the male, are responsible for the pair-bonding behavior in prairie voles [81]. The pattern of oxytocin and vasopressin immunoreactivity in the brains of the two species does not differ substantially, but the distribution of their receptors does [82,83•]. Furthermore, the gene for the vasopressin receptor differs in its 5'-flanking region but not in the coding region [84**]. This difference may determine which regions of the brain express the receptor. A transgenic mouse expressing the prairie vole vasopressin receptor gene shows a pattern of vasopressin receptor expression that is similar to that seen in the prairie vole, and increased affiliative behavior in response to vasopressin, also reminiscent of prairie voles [84..]. This work demonstrates that the localization of receptors that underlie differences in behavior can be accomplished easily through mutations in the promoter regions of particular genes.

Importance of knowing phylogeny

When comparing the neuronal circuitry underlying behaviors in two species, it is important to understand the phylogenetic relationships between the species and to conduct an out-group comparison. For example, by comparing the neuronal responses to serotonin with a phylogeny based on other characters, it was shown that the lack of a sensory neuron excitability response in the mollusc Dolabrifera was attributable to a secondary loss of the response after splitting from other groups that retained it [79]. Another good example is work on the jamming avoidance response (JAR) in weakly electric fish. Electroreception arose early in vertebrate evolution and was subsequently lost and 're-evolved' numerous times [85]. Two different genera of wave-type electric fish both evolved a JAR to prevent their electric signals from being confused with similar signals from nearby fish [86,87]. This behavior has very particular requirements, and thus both species use an identical set of computational rules to perform the task. However, the circuitry underlying these apparently identical behaviors resides in different brain areas, revealing that the two behaviors evolved independently [88].

Conclusions

Although nervous systems tend to be more evolutionarily conserved than other parts of the body, centrally generated behavioral patterns do change. It appears that rather subtle changes in the nervous system can cause large and important changes in the behavior of an organism. This fits very well with our understanding of how neuronal circuits and even single neurons can show dramatic alterations in activity with very small changes in parameters such as the density of ion channels [89]. The modulatory inputs to neural networks seem to be a very plastic trait in CNS evolution. A change in the distribution of receptors or in the expression of peptide co-transmitters can be made very easily through changes in the promoter regions of the genes. The nervous system is organized in such a way that it can accept these changes and incorporate them to generate a novel species-specific behavior.

Acknowledgements

We thank Sarah Pallas, Donald Edwards, Ronald Hoy, Bruce Johnson, and Charles Derby for helpful suggestions on this manuscript. PS Katz's work is supported by National Institutes of Health (NIH) grant NS-35371. RM Harris-Warrick's work is supported by NIH grants NS17323 and NS35631.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Crews D: Species diversity and the evolution of behavioral controlling mechanisms. Ann NY Acad Sci 1997, 807:1-21.
- 2. Lingle S: Escape gaits of white-tailed deer, mule deer and their hybrids: gaits observed and patterns of limb coordination. *Behaviour* 1992, **122**:153-181.
- 3. Ryan MJ: Sexual selection, receiver biases, and the evolution of sex differences. *Science* 1998, **281**:1999-2003.

- 4. Ritchie MG: The shape of female mating preferences. Proc Natl Acad Sci USA 1996, 93:14628-14631.
- Balaban E: Changes in multiple brain regions underlie species 5. differences in a complex, congenital behavior. Proc Natl Acad Sci USA 1997, 94:2001-2006. ..

Chickens (Gallus gallus domesticus) and Japanese quail (Coturnix coturnix japonica) exhibit species-specific vocalizations that include complex head movements. It was found that transplanting regions of the midbrain or brainstem from the embryo of one species to the other could transform the adult behavior of the host species, causing it to express components of the behavior exhibited by the donor species. These results are interpreted to mean that species differences in behavior are attributable to localized changes in the neuronal circuitry.

6. Faulkes Z, Paul DH: Coordination between the legs and tail during digging and swimming in sand crabs. J Comp Physiol [A] 1997, 180:161-169.

Suggests that digging behavior in sand crabs may have originated as a modification of the walking motor pattern. Reports that one group of sand crabs has evolved a novel form of swimming, allowing a differential coordination of its limbs. This indicates that progressive changes in behavior can occur in particular lineages of species.

- 7. Faulkes Z, Paul DH: Digging in sand crabs (Decapoda, Anomura, Hippoidea): interleg coordination. J Exp Biol 1997, 200:793-805.
- Gatesy SM, Dial KP: Locomotor modules and the evolution of 8. avian flight. Evolution 1996, 50:331-340.
- Wainwright PC, Turingan RG: Evolution of pufferfish inflation 9

behavior. Evolution 1997, 51:506-518.

Pufferfish have evolved a behavioral innovation that is reflected in their anatomical organization. This paper uses the phylogeny of tetrodontiform fishes to derive a hypothesis for the origin of the inflation behavior. The study convincingly demonstrates that this novel behavior evolved from a pre-existing 'coughing' behavior that was refined into a water-blowing behavior in the lineage leading to pufferfish.

- 10. Van Staaden MJ, Römer H: Evolutionary transition from stretch to hearing organs in ancient grasshoppers. Nature 1998, 394:773-776.
- 11. Paul DH, Wilson LJ: Replacement of an inherited stretch receptor by a newly evolved stretch receptor in hippid sand crabs. J Comp Neurol 1994, **350**:150-160.
- 12. Arbas EA, Meinertzhagen IA, Shaw SR: Evolution in nervous systems. Annu Rev Neurosci 1991, 14:9-38.
- 13. Tierney AJ: Evolutionary implications of neural circuit structure and function. Behav Processes 1995. 35:173-182.
- 14. Katz PS: Neuromodulation and the evolution of a simple motor system. Semin Neurosci 1991, 3:379-389.
- 15. Katz PS, Tazaki K: Comparative and evolutionary aspects of the crustacean stomatogastric system. In Dynamic Biological Networks: The Stomatogastric Nervous System. Edited by Harris-Warrick RM, Marder E, Šelverston AI, Moulins M. Cambridge, Massachusetts: MIT Press; 1992:221-261.
- 16. Friel JP, Wainwright PC: Evolution of complexity in motor patterns and jaw musculature of tetraodontiform fishes. J Exp Biol 1999, 202:867-880.
- Friel JP, Wainwright PC: Evolution of motor patterns in 17.
- tetraodontiform fishes: does muscle duplication lead to functional diversification? Brain Behav Evol 1998, 52:159-170.

Duplication of jaw muscles has occurred several times within the teleost fish order Tetraodontiformes. This study provides evidence that such increases in the complexity of the peripheral apparatus led to increasing complexity of the motor program of the species in that lineage.

- 18. Kavanau JL: Conservative behavioral evolution, the neural substrate. Anim Behav 1990, 39:758-767.
- 19. Smith KK: Are neuromotor systems conserved in evolution? Brain Behav Evol 1994, 43:293-305.
- 20. Finlay BL, Pallas SL: Control of cell number in the developing mammalian visual system. Prog Neurobiol 1989, 32:207-234.
- 21. Caldero J, Prevette D, Mei X, Oakley RA, Li L, Milligan C, Houenou L, Burek M, Oppenheim RW: Peripheral target regulation of the development and survival of spinal sensory and motor neurons in the chick embryo. J Neurosci 1998, 18:356-370.
- 22. Wang Y, Smallwood PM, Cowan M, Blesh D, Lawler A, Nathans J:
- Mutually exclusive expression of human red and green visual pigment-reporter transgenes occurs at high frequency in murine

cone photoreceptors. Proc Natl Acad Sci USA 1999, 96:5251-5256.

This paper deals with the mechanisms that allow peripheral changes to be incorporated into functional neuronal circuits. Only humans and some other old world primates have trichromatic color vision; other mammals have dichromatic color vision. The green (middle-wavelength) pigment arose from a duplication of the red (long-wavelength) pigment gene on the X chromo-some in the primate lineage about 30–40 million years ago. The authors asked how trichromatic color vision could arise from a circuit that has two types of photoreceptors. They did this by creating transgenic mice that carry a single copy of the human X chromosome visual pigment gene array in which the genes for the visual pigments were swapped for reporter genes that can be differentially stained. They found that the two gene products were differentially expressed in photoreceptors. Thus, a simple gene-duplication event could have given rise to different populations of photoreceptors. The nature of the retinal circuitry would automatically create color opponent center-surround ganglion cells, and Hebbian-type mechanisms could then allow for central sorting of the ganglion cells.

- Edwards JS: The evolution of insect flight: implications for the 23. evolution of the nervous system. Brain Behav Evol 1997, 50:8-12.
- Kempf SC, Page LR, Pires A: Development of serotonin-like 24. immunoreactivity in the embryos and larvae of nudibranch mollusks with emphasis on the structure and possible function of the apical sensory organ. J Comp Neurol 1997, 386:507-528.

This paper suggests that the apical sensory organ found in the larva of many invertebrate phyla, including molluscs and annelids, might be homologous and contain homologous neurons in these diverse groups. Certainly within the molluscs, five serotonergic neurons with similar morphology are found in the larva of species with highly divergent adult forms.

- 25. Buschbeck EK, Strausfeld NJ: Visual motion-detection circuits in flies: small-field retinotopic elements responding to motion are evolutionarily conserved across taxa. J Neurosci 1996, 16:4563-4578
- Sudlow LC, Jing J, Moroz LL, Gillette R: Serotonin immunoreactivity 26. in the central nervous system of the marine molluscs Pleurobranchaea californica and Tritonia diomedea. J Comp Neurol 1998, 395:466-480.
- 27. Beltz BS: Distribution and functional anatomy of amine-containing neurons in decapod crustaceans. Microsc Res Tech 1999, 44:105-120.
- 28. Harrison PJ, MacMillan DL, Young HM: Serotonin immunoreactivity in the ventral nerve cord of the primitive crustacean Anaspides tasmaniae closely resembles that of crayfish. J Exp Biol 1995, 198:531-535.
- 29. Paul DH: Pedigrees of neurobehavioral circuits: tracing the evolution of novel behaviors by comparing motor patterns, muscles, and neurons in members of related taxa. Brain Behav Evol 1991. 38:226-239.
- 30. Finlay BL, Hersman MN, Darlington RB: Patterns of vertebrate neurogenesis and the paths of vertebrate evolution. Brain Behav Evol 1998, 52:232-242.
- 31. Truman JW, Ball EE: Patterns of embryonic neurogenesis in a primitive wingless insect, the silverfish, Ctenolepisma longicaudata: comparison with those seen in flying insects. Dev Genes Evol 1998, 208:357-368.

The authors report that the number of neuroblasts remains constant across all insect orders. The final number of neurons is determined by the relative proliferation and subsequent cell death within each neuronal lineage

- 32. Williams RW, Cavada C, Reinoso-Suárez F: Rapid evolution of the visual system: a cellular assay of the retina and dorsal lateral geniculate nucleus of the Spanish wildcat and the domestic cat. J Neurosci 1993, 13:208-228.
- Volman SF, Grubb TC Jr, Schuett KC: Relative hippocampal volume in relation to food-storing behavior in four species of woodpeckers. Brain Behav Evol 1997, 49:110-120.
- 34. Witten JL, Truman JW: Distribution of GABA-like immunoreactive neurons in insects suggests lineage homology. J Comp Neurol 1998, **398**:515-528.

The authors identified similar groups of GABAergic neurons in insects and crustaceans. The number of cells in each of the neuronal groupings varies between insect orders. Thus, although homologous neurons are found, their numbers differ significantly.

35. Finger TE: Feeding patterns and brain evolution in ostariophysean fishes. Acta Physiol Scand 1997, 161:59-66.

Reports that goldfish have diverged from most other teleost fishes in the organization of their vagal taste nucleus. Although the synaptic connectivity of this nucleus is the same as in other teleosts, in goldfish it is a laminated structure that allows a precise mapping of sensory and motor components. Furthermore, the laminar organization provides a developmental framework that more easily permits addition of neurons to increase computational capacity.

- Finlay BL, Darlington RB: Linked regularities in the development and evolution of mammalian brains. *Science* 1995, 268:1578-1584.
- 37. Kiehn O, Hounsgaard J, Sillar KT: Basic building blocks of
- vertebrate spinal central pattern generators. In Neurons, Networks and Motor Behavior. Edited by Stein PSG, Grillner S, Selverston A, Stuart DG. Cambridge, Massachusetts: MIT Press; 1997:47-59.

This review summarizes the common building blocks of the central pattern generators for locomotion in the spinal cords of a number of species, including cat, rat, lamprey and *Xenopus*. These networks contain unit burst generators on the left and right sides of the spinal cord that are coupled primarily by crossed glycinergic inhibitory and weaker excitatory connections. Rhythm generation depends critically on glutamatergic activation of NMDA and non-NMDA receptors. Nonlinear response properties of the interneurons, including postinhibitory rebound, plateau potentials and intrinsic bursting, are important in shaping the motor pattern and are under control of modulatory inputs, including serotonin, noradrenaline, and several peptides. Thus, the basic structure of the locomotor pattern generator is conserved between classes within the chordate phylum.

- 38. Soffe SR, Perrins R: Neuronal firing properties and swimming
- motor patterns in young tadpoles of four amphibians: Xenopus, Rana, Bufo and Triturus. J Comp Physiol [A] 1997, 181:71-81.

Reports that equivalent motor neurons in four amphibian species (belonging to the genera *Xenopus, Rana, Bufo* and *Triturus*) differ in their firing patterns during locomotion. Differences in motor neuron properties exist, but are not sufficient to account for the different activity patterns. The authors suggest that the synaptic input to the motor neurons exhibits species differences. However, the basis for this difference was not determined in this study.

- Perrins R, Soffe SR: Composition of the excitatory drive during swimming in two amphibian embryos: Rana and Bufo. J Comp Physiol [A] 1996, 179:563-573.
- Jing J, Gillette R: Central pattern generator for escape swimming
 in the notaspid sea slug *Pleurobranchaea californica*. J Neurophysiol 1999, 81:654-667.

The authors identified many of the neurons comprising the central pattern generator (CPG) circuit for swimming in *Pleurobranchaea*. Based on their neurotransmitter phenotypes, location, synaptic actions, and activity pattern during the swimming motor program, it was judged that most of these neurons are homologous to neurons in the swim CPG of the distantly related nudibranch mollusc *Tritonia diomedea*. Some neurons were found in one species, but not in the other. This is probably attributable to incomplete characterization of both circuits. The strong similarities indicate a general conservation of the components of the circuit. Differences were seen in the strength of particular synapses and electric connections.

- 41. Tazaki K, Tazaki Y: Neural control of the pyloric region in the
- foregut of the shrimp Penaeus (Decapoda: Penaeidae). J Comp Physiol A 1997, 181:367-382.

This paper describes the structure of the pyloric network in a primitive decapod, *Penaeus*. All of the neurons that are present in more advanced decapods are also present in this shrimp, and the basic synaptic circuitry is virtually identical. A detailed comparison with three more advanced species is given.

- Tazaki K, Chiba C: Cellular properties and modulation of the stomatogastric ganglion neurons of a stomatopod, Squilla oratoria. J Comp Physiol [A] 1993, 173:85-101.
- Tazaki K: Motor pattern generation of the posterior cardiac plate pyloric system in the stomatogastric ganglion of the mantis shrimp Squilla oratoria. J Comp Physiol [A] 1993, 172:369-387.
- 44. Meyrand P, Moulins M: Phylogenetic plasticity of crustacean stomatogastric circuits. I. Pyloric patterns and pyloric circuit of the shrimp *Palaemon serratus. J Exp Biol* 1988, **138**:107-132.
- 45. Combes D, Meyrand P, Simmers J: Motor pattern specification by
 dual descending pathways to a lobster rhythm-generating network. J Neurosci 1999, 19:3610-3619.

The gastric mill central pattern generator circuit in the lobster, *Homarus* gammarus, is essentially identical to the circuits in spiny lobsters and crabs in the identity of cells and their synaptic interconnectivity. There are changes such as differences in the number of a particular type of motor neuron and the strength of some connections. Despite these differences, the output of the circuits under comparable conditions is remarkably similar.

 Meyrand P, Moulins M: Phylogenetic plasticity of crustacean stomatogastric circuits. II. Extrinsic inputs to the pyloric circuit of the shrimp Palaemon serratus. J Exp Biol 1988, 138:133-153.

- Tazaki K, Chiba C: Glutamate, acetylcholine, and γ-aminobutyric acid as transmitters in the pyloric system of the stomatogastric ganglion of a stomatopod, Squilla oratoria. J Comp Physiol [A] 1994, 175:487-504.
- Green CS, Soffe SR: Roles of ascending inhibition during two rhythmic motor patterns in *Xenopus* tadpoles. J Neurophysiol 1998, 79:2316-2328.
- Soffe SR: Motor patterns for two distinct rhythmic behaviors evoked by excitatory amino acid agonists in the Xenopus embryo spinal cord. J Neurophysiol 1996, 75:1815-1825.
- Soffe SR: The pattern of sensory discharge can determine the motor response in young *Xenopus* tadpoles. *J Comp Physiol [A]* 1997, 180:711-715.
- 51. Clemens S, Meyrand P, Simmers J: Feeding-induced changes in temporal patterning of muscle activity in the lobster stomatogastric system. *Neurosci Lett* 1998, **254**:65-68.
- Kiehn O, Katz PS: Making circuits dance: neuromodulation of motor systems. In Beyond Neurotransmission: Neuromodulation and its Importance for Information Processing. Edited by Katz PS. Oxford: Oxford University Press; 1999:275-317.
- Harris-Warrick RM, Nagy F, Nusbaum MP: Neuromodulation of stomatogastric networks by identified neurons and transmitters. In Dynamic Biological Networks: The Stomatogastric Nervous System. Edited by Harris-Warrick RM, Marder E, Selverston AI, Moulins M. Cambridge, Massachussetts: MIT Press; 1992:87-137.
- Casasnovas B, Meyrand P: Functional differentiation of adult neural circuits from a single embryonic network. J Neurosci 1995, 15:5703-5718.
- Fénelon VS, Casasnovas B, Simmers J, Meyrand P: Development of rhythmic pattern generators. Curr Opin Neurobiol 1998, 8:705-709.
- Fénelon VS, Casasnovas B, Faumont S, Meyrand P: Ontogenetic alteration in peptidergic expression within a stable neuronal population in lobster stomatogastric nervous system. *J Comp Neurol* 1998, 399:289-305.
- Bekoff A: Neuroethological approaches to the study of motor development in chicks: achievements and challenges. J Neurobiol 1992, 23:1486-1505.
- Ayali A, Harris-Warrick RM: Monoamine control of the pacemaker kernel and cycle frequency in the lobster pyloric network. *J Neurosci* 1999, 19:6712-6722.
- 59. Soffe SR: Two distinct rhythmic motor patterns are driven by common premotor and motor neurons in a simple vertebrate spinal cord. *J Neurosci* 1993, **13**:4456-4469.
- Meyrand P, Simmers J, Moulins M: Dynamic construction of a neural network from multiple pattern generators in the lobster stomatogastric nervous system. J Neurosci 1994, 14:630-644.
- 61. Stein PS, McCullough ML, Currie SN: Spinal motor patterns in the turtle. Ann NY Acad Sci 1998, 860:142-154.
- 62. Paggett KC, Gupta V, McClellan AD: Adaptive variations of undulatory behaviors in larval lamprey: comparison of swimming and burrowing. *Exp Brain Res* 1998, **119**:213-223.
- 63. Dickinson PS: Interactions among neural networks for behavior. Curr Opin Neurobiol 1995, 5:792-798.
- Heiligenberg W, Metzner W, Wong CJH, Keller CH: Motor control of the jamming avoidance response of *Apteronotus leptorhynchus*: evolutionary changes of a behavior and its neuronal substrates. *J Comp Physiol* [A] 1996, **179**:653-674.
- 65. Juranek J, Metzner W: Cellular characterization of synaptic
 modulations of a neuronal oscillator in electric fish. J Comp Physiol [A] 1997, 181:393-414.

Reports that the inputs to the pacemaker nucleus that are common to both *Eigenmannia* and *Apteronotus* produce species-specific modulatory actions on relay cells, contributing to differences in the jamming avoidance responses of animals in these genera.

- Sillar KT, Woolston A-M, Wedderburn JFS: Involvement of brainstem serotonergic interneurons in the development of a vertebrate spinal locomotor circuit. Proc R Soc Lond [Biol] 1995, 259:65-70.
- Scrymgeour-Wedderburn JF, Reith CA, Sillar KT: Voltage oscillations in Xenopus spinal cord neurons: developmental onset and dependence on co-activation of NMDA and 5HT receptors. Eur J Neurosci 1997, 9:1473-1482.

- Woolston A-M, Wedderburn JFS, Sillar KT: Descending serotonergic spinal projections and modulation of locomotor rhythmicity in *Rana temporaria* embryos. *Proc R Soc Lond [Biol]* 1994, 255:73-79.
- 69. Nelson TE, King JS, Bishop GA: Distribution of tyrosine hydroxylase-immunoreactive afferents to the cerebellum differs between species. J Comp Neurol 1997, **379**:443-454.
- 70. Skiebe P: Allatostatin-like immunoreactivity in the stomatogastric
- nervous system and the pericardial organs of the crab Cancer pagurus, the lobster Homarus americanus, and the crayfish Cherax destructor and Procambarus clarkii. J Comp Neurol 1999, 403:85-105.

This study looked at innervation of the stomatogastric ganglion (STG) by fibers displaying allatostatin (AST)-like immunoreactivity in four decapod crustaceans. The STGs of the four species had different sources of AST-like immunoreactivity: all of them received input from the sensory gastropyloric receptor (GPR) neurons, but three of them also received descending AST-like inputs from other ganglia, and two species had AST-labeled neurons within the STG itself. This study shows how variable the pattern of peptide immunoreactivity can be among closely related species.

- Turrigiano GG, Selverston Al: Distribution of cholecystokinin-like immunoreactivity within the stomatogastric nervous systems of four species of decapod Crustacea. J Comp Neurol 1991, 305:164-176.
- Tierney AJ, Blanck J, Mercier AJ: FMRFamide-like peptides in the crayfish (*Procambarus clarkii*) stomatogastric nervous system: distribution and effects on the pyloric motor pattern. *J Exp Biol* 1997, 200:3221-3233.
- Tierney AJ, Godleski MS, Rattananont P: Serotonin-like immunoreactivity in the stomatogastric nervous systems of crayfishes from four genera. *Cell Tissue Res* 1999, 295:537-551.
- Beltz B, Eisen JS, Flamm RE, Harris-Warrick RM, Hooper SL, Marder E: Serotonergic innervation and modulation of the stomatogastric ganglion of three decapod crustaceans (*Panulirus interruptus*, *Homarus americanus* and *Cancer irroratus*). J Exp Biol 1984, **109**:35-54.
- Katz PS, Harris-Warrick RM: Actions of identified neuromodulatory neurons in a simple motor system. *Trends Neurosci* 1990, 13:367-373.
- Turrigiano GG, Selverston Al: A cholecystokinin-like hormone activates a feeding-related neural circuit in lobster. *Nature* 1990, 344:866-868.
- 77. Blitz DM, Christie AE, Coleman MJ, Norris BJ, Marder E,
- Nusbaum MP: Different proctolin neurons elicit distinct motor patterns from a multifunctional neuronal network. J Neurosci 1999, 19:5449-5463.

Reports that three modulatory neurons to the crab stomatogastric ganglion all contain the peptide proctolin, yet they each elicit distinct motor patterns. These differences seem to arise in part from different co-transmitter complements in the three neurons, different target neurons, different influences on common target neurons, different abilities to recruit additional modulatory neurons to fire simultaneously, and possibly different amounts of proctolin released from nerve terminals. Thus, the action of a neuromodulator can be very different if it is released from a different neuron.

 Nässel DR, Eckert M, Muren JE, Penzlin H: Species-specific action and distribution of tachykinin-related peptides in the foregut of the cockroaches *Leucophaea maderae* and *Periplaneta americana. J Exp Biol* 1998, 201:1615-1626.

- Wright WG, Kirschman D, Rozen D, Maynard B: Phylogenetic analysis of learning-related neuromodulation in molluscan mechanosensory neurons. *Evolution* 1996, 50:2248-2263.
- Wright WG: Evolution of nonassociative learning: behavioral analysis of a phylogenetic lesion. Neurobiol Learn Mem 1998, 69:326-337.
- Young LJ, Wang ZX, Insel TR: Neuroendocrine bases of monogamy. Trends Neurosci 1998, 21:71-75.
- Insel TR, Winslow JT, Wang ZX, Young L, Hulihan TJ: Oxytocin and the molecular basis of monogamy. *Adv Exp Med Biol* 1995, 395:227-234.
- 83. Young LJ, Winslow JT, Nilsen R, Insel TR: Species differences in V1a
 receptor gene expression in monogamous and nonmonogamous

voles: behavioral consequences. Behav Neurosci 1997, 111:599-605. The prairie vole is monogamous and exhibits extensive parental care, whereas the montane vole mates promiscuously and is asocial. The authors observed significant differences in the distribution of the V_{1a} vasopressin receptor between the two species using *in situ* hybridization for receptor RNA and receptor autoradiography for the receptor protein. This finding correlates with distinct species-dependent differences in behavioral responses to central administered arginine-vasopressin. These data suggest that phylogenetic plasticity in vasopressin receptor localization may play a role in the evolution of social behavior.

 Young LJ, Nilsen R, Waymire KG, Macgregor GR, Insel TR: Increased
 affiliative response to vasopressin in mice expressing the V_{1a} receptor from a monogamous vole. *Nature* 1999, 400:766-768.

Centrally administered arginine-vasopressin increases affiliative behavior towards a female in the monogamous prairie vole but not the asocial montane vole. The authors cloned the V1a receptor gene from the two social species and two asocial species of voles. While the coding region was nearly identical, the 5'-flanking region differed significantly. The two species of affiliative prairie voles contain a 428 bp sequence (rich in microsatellite DNA sequences) that is lacking in the promoter region of two asocial species of montane vole. Transgenic mice were created with the prairie vole V1a receptor minigene, including 2.2 kb of the 5'-flanking region. This gene was expressed in the same locations as in the prairie vole but not the montane vole. In addition, the transgenic mice showed increased affiliative behavior towards females upon central administration of arginine-vasopressin; this behavior was not seen in non-transgenic littermates. These results suggest that changes in the distribution of the V1a receptor, attributable to changes in the 5'-flanking sequence of the gene, can lead to marked changes in social behavior between species.

- New JG: The evolution of vertebrate electrosensory systems. Brain Behav Evol 1997, 50:244-252.
- Metzner W: Neural circuitry for communication and jamming avoidance in gymnotiform electric fish. J Exp Biol 1999, 202:1365-1375.
- Kawasaki M: Sensory hyperacuity in the jamming avoidance response of weakly electric fish. Curr Opin Neurobiol 1997, 7:473-479.
- Kawasaki M: Comparative analysis of the jamming avoidance response in African and South American wave-type electric fishes. *Biol Bull* 1996, **191**:103-108.
- Turrigiano GG: Message received: cellular responses to neuromodulatory signals. In Beyond Neurotransmission: Neuromodulation and its Importance for Information Processing. Edited by Katz PS. Oxford: Oxford University Press; 1999:121-159.